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Factor Xa inhibition with apixaban does not influence cardiac remodeling in rats with heart failure after myocardial infarction

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ABSTRACT

Background: Heart failure (HF) is considered to be a prothrombotic condition and it has been suggested that coagulation factors contribute to maladaptive cardiac remodelling via activation of the protease-activated receptor 1 (PAR1). We tested the hypothesis that anticoagulation with the factor Xa (FXa) inhibitor apixaban would ameliorate cardiac remodelling in rats with HF after myocardial infarction (MI).

Methods and results: Male Sprague-Dawley rats were either subjected to permanent ligation of the left ascending coronary artery (MI) or sham surgery. The MI and sham animals were randomly allocated to treatment with placebo or apixaban in the chow (150 mg/kg/day), starting 2 weeks after surgery. Cardiac function was assessed using echocardiography and histological and molecular markers of cardiac hypertrophy were assessed in the left ventricle (LV).

Apixaban resulted in a fivefold increase in anti-FXa activity compared to vehicle, but no overt bleeding was observed and haematocrit levels remained similar in apixaban- and vehicle- treated groups. After 10 weeks of treatment, LV ejection fraction was 42 ± 3 % in the MI group treated with apixaban and 37 ± 2 in the vehicle treated MI group ($p > 0.05$). Both vehicle and apixaban-treated MI groups also displayed similar degrees of LV dilatation, LV hypertrophy and interstitial fibrosis. Histological and molecular markers for pathological remodelling were also comparable between groups, as was the activity of signalling pathways downstream of the PAR1 receptor.

Conclusion: FXa inhibition with apixaban does not influence pathological cardiac remodelling after MI. These data do not support the use of FXa inhibitor in HF patients with the aim to amend the severity of HF.

Keywords: Anticoagulant • Heart failure • Cardiac function • Cardiac remodelling

INTRODUCTION

Heart failure (HF) is major global health problem that is reaching epidemic proportions in the near future [1,2]. Despite the large range of pharmacological and device-based therapies available, mortality and morbidity remain high [3]. Patients with HF are at increased risk of stroke and other thromboembolic events and are also more likely to succumb from these events [4–9]. The higher incidence of thromboembolic events is observed both in patients with sinus rhythm with atrial fibrillation [10,11], suggesting that HF should be considered as a prothrombotic or hypercoagulable state. Interestingly, coagulation factors such as thrombin and FXa can exert direct effects on the heart, which are thought to promote inflammation, endothelial dysfunction and maladaptive cardiac remodelling [12]. Anticoagulants could therefore offer therapeutic benefits in HF patients beyond prevention of thromboembolic events [10].

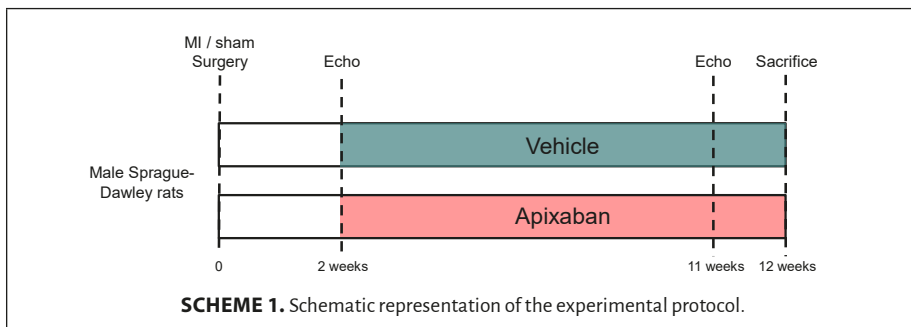
The effects of thrombin and FXa on myocardial tissues are thought to be governed by protease-activated receptors (PARs), which coordinate a myriad of cellular responses in multiple cell types. PAR1 and PAR2 are expressed in cardiac tissue and it has been proposed that these receptors contribute to the progression of HF [13–15]. Indeed, cardiomyocyte-specific overexpression of PAR1 induces cardiac hypertrophy that rapidly progressed into dilated cardiomyopathy [16]. Conversely, deletion of the PAR1 receptor attenuates cardiac remodelling after a myocardial infarction (MI) in mice [16]. Nevertheless, direct evidence for enhanced activity of PARs in HF is sparse and the exact role of PAR signalling in cardiac remodelling remains poorly defined. Furthermore, it is unknown whether PAR receptor activation in HF is amendable by anticoagulant therapy. In this study, we tested the hypothesis that inhibition of PAR signalling by the direct FXa inhibitor apixaban could attenuate cardiac remodelling in rats with established LV dysfunction after MI.

METHODS

Experimental protocol

Male Sprague Dawley rats (Envigo, The Netherlands) were randomized to treatment with chow containing apixaban or control chow, starting 2 weeks after MI surgery. We chose this protocol to represent a population of stable chronic HF after a large myocardial infarction, which still represents the majority of patients with chronic HF. Here, we examined the effects of clinically relevant doses of apixaban on cardiac remodeling in rats with HF after MI [17]. Treatment allocation was stratified according to left ventricular ejection fraction (LVEF) to ensure that the baseline cardiac function is similar in the apixaban and the vehicle groups. After 10 weeks of treatment, rats were anesthetized, blood was drawn and the hearts

were rapidly excised for further analysis. Rats with an infarct size of less than 15% were excluded from analysis as these small infarcts are haemodynamically fully compensated [18]. The experimental protocol is illustrated in Scheme 1.



Ethical statement

The experimental protocol was approved by the Animal Ethical Committee of University of Groningen (IvD number: 16487-02-001). The investigation conforms to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication no. 85-23, revised 1996). We followed ARRIVE guidelines when reporting this study.

MI surgery

Rats were randomized to HF or sham surgery under isoflurane (2.5%) inhalation anaesthesia. After left-sided thoracotomy, HF was induced by permanent ligating of the proximal portion of the left coronary artery as previously described [19]. Sham operated rats underwent the same procedure but without coronary ligation.

Investigational drug

Apixaban was kindly supplied by Bristol-Myers Squibb (BMS, USA). Apixaban was mixed with standard rat chow (R/M-H V1534-70, Ssniff, Germany) in a final concentration 1,95 gram/kg intended to reach an average dose of 150 mg/kg. Standard rat chow (R/M-H V1534-70, Ssniff, Germany) was used as control (vehicle).

Echocardiography

Two weeks after surgery and 1 week before termination, the M-mode and 2D echocardiography was performed using a Vivid 7 echo machine (GE Healthcare) equipped with a 10-MHz phase array linear transducer for serially assessment of cardiac structure and function as previously described [19].

Invasive haemodynamic measurements

Prior to sacrifice, invasive hemodynamics were analysed by aortic and LV catheterization as previously described [19]. The right carotid artery was isolated, punctured, and a 1.9 F rat pressure-volume catheter (Scisense, London, Ontario, Canada) was inserted into the right carotid artery. The tip of the catheter was advanced through the aorta into the LV cavity. Heart rate (HR), left ventricular end-systolic (LVESP) and end-diastolic (LVEDP) pressures, and maximal rates of increase and decrease in developed LV pressure (dP/dt_{max} and dP/dt_{min}) were determined. The data were acquired using a PowerLab data acquisition system (ADInstruments, Colorado Springs, CO) and analysed with a LabChart 8 software.

Infarct size, cardiomyocyte size and interstitial fibrosis measurement

Rats were euthanized under isoflurane anaesthesia. Heart was rapidly excised and weighed. The mid-papillary slice of the LV was fixed in 4% formaldehyde and paraffin-embedded. The infarct size was calculated using midline length methods as percentage of the scar length to the total LV circumference on Masson's trichrome-stained section, as described previously [19,20]. Furthermore, Masson's trichrome staining was also used to evaluate the extent of interstitial fibrosis. Hamamatsu microscope was used to capture the whole tissue section and Aperio ImageScope software was used to quantify fibrosis in the non-infarcted LV [19]. Finally, sections were stained with FITC Labelled wheat germ agglutinin (WGA) to determine cardiomyocyte size as previously described [19]. Cell size from transversally cut cardiomyocytes in the non-infarcted LV was measured using image analyser (Zeiss KS400, Germany) and quantified using ImageJ software (National Institutes of Health, Bethesda, MD, USA). The investigators analysing the data were blinded to the treatment allocation.

Blood and urine measurements

Blood samples were obtained via a tail vein under isoflurane anaesthesia to determine anti-factor Xa activity (AXA), pro-thrombin time (PT) and activated partial thromboplastin time (APTT) levels. At sacrifice, 8 ml of blood was drawn from the abdominal aorta (either anti-coagulated with sodium citrate or EDTA), and urine was collected directly from bladder. Complete blood count was determined on the day of sacrifice using a Sysmex hematology analyzer (Sysmex XN-10, Sysmex Corporation, Japan).

Prothrombin time (PT), activated partial thromboplastin time (APTT) and anti-factor Xa activity (AXA) assay

To determine the effect of apixaban on PT and APTT levels, each sample was tested using Innovin (PT) and Actin FS (APTT) reagents (Siemens Healthcare Diagnostics). Plasma apixaban concentration was assessed using a chromogenic anti-factor Xa activity (AXA) assay, the Berichrom Heparin Assay (Dade Behring, Marburg, Germany) as this is the most reliable method to measure the pharmacodynamics of apixaban [21].

Urine occult blood test

Haemoglobin/red blood cells was determined by a semiquantitative method using Combur10 Test Sticks (Roche Diagnostics GmbH, Mannheim, Germany).

Quantitative real-time PCR

RNA was extracted from the non-infarcted LV using TRIzol reagent (Invitrogen Corp., Carlsbad, CA, USA), as previously described [19,22] and the NanoDrop device was used to measure RNA concentration. Random primer mix was used to prepare first-stranded DNA and thereafter used as a template for quantitative real-time reverse-transcriptase-PCR (qRT-PCR) (25 ng/reaction). mRNA levels obtained by a qRT-PCR using C1000 Thermal Cycler CFX384 Real-Time PCR Detection System (Bio-Rad Laboratories, Veenendaal, The Netherlands). 36B4 reference gene was used to correct all measured mRNA expression. Primer sequences can be found in supplementary *Table S1*.

Western blot

Frozen non-infarcted LV tissue was homogenized in ice-cold lysis buffer containing phosphatase inhibitor cocktail 1 (Sigma) and protease inhibitor (ROCHE) as described previously [19]. Bio-Rad DC Protein Assay (Bio-Rad Laboratories, Veenendaal, The Netherlands) was used for protein quantifications with bovine albumin as a standard, as described before [19]. Immunoblotting was performed using primary antibodies from commercial suppliers (Supplementary *Table S2*). Immunoblots were incubated with appropriate secondary antibodies for 1 h at room temperature. Signals were detected by ECL (ParkinElmer, Waltham, MA, USA). Blots were quantified using ImageJ software (National Institutes of Health, Bethesda, MD, USA). The density of each band was normalized to GAPDH acting as a loading control and presented as fold change over Sham-veh group.

RhoA activity assay

RhoA activity was measured according to manufacturer's protocol (BK124; Cytoskeleton Inc.), as previously described [23].

Statistical analysis

Data are presented as means \pm standard errors of the mean (SEM). To compare normally distributed parameters, one-way analysis of variance (ANOVA) followed by Tukey's Post Hoc test was used. When data were not normally distributed, a non-parametric Kruskal-Wallis test followed by a Mann-Whitney U test with correction for multiple comparisons was used. Wilcoxon signed rank test was used to evaluate LVEF post-MI vs. before termination. Differences were considered significant at $p < 0.05$. IBM SPSS Statistics for Windows, Version 23.0 (IBM Corp, USA) was used to perform all statistical analysis.

RESULTS

We performed MI or sham surgery on a total of 78 male Sprague Dawley rats; 21 rats (27%) died during the surgical procedure, all remaining rats survived the rest of the study. A total of 4 rats with an infarct <15% of the LV were excluded from further analysis, leaving a total of 53 rats for the analysis. The final group sizes were 8 for the sham-apixaban group, 13 for the sham-vehicle group, 17 for the MI-apixaban group and 15 for the MI-vehicle group.

TABLE 1. General characteristics and haematological parameters.

Parameters	Sham-vehicle	Sham-apixaban	MI-vehicle	MI-apixaban
Food intake (g/day)	31.91 ± 0.5	29.87 ± 0.7	30.27 ± 0.5	29.64 ± 0.3
Water intake (ml/day)	31.97 ± 0.9	29.86 ± 0.6	30.89 ± 1.0	28.50 ± 0.4
Body weight change (g)	91.15 ± 3.2	93.25 ± 4.8	94.73 ± 4.8	93.24 ± 4.8
AXA (ng/ml)	21 ± 2	109 ± 17†	19 ± 1	103 ± 11#
PT (sec)	11.12 ± 0.12	11.00 ± 0.10	11.31 ± 0.21	11.17 ± 0.17
APTT (sec)	15.48 ± 1.13	16.74 ± 0.62	17.24 ± 1.20	16.91 ± 0.94
WBC (10 ⁹ /L)	9.34 ± 2.18	11.34 ± 0.68	11.23 ± 1.24	10.66 ± 0.80
RBC (10 ⁹ /L)	9.01 ± 0.17	8.94 ± 0.10	8.99 ± 0.10	9.32 ± 0.23
HCT (mmol/L)	10.02 ± 0.25	9.84 ± 0.12	9.79 ± 0.13	10.00 ± 0.12
HGB (L/L)	0.503 ± 0.02	0.489 ± 0.01	0.493 ± 0.01	0.505 ± 0.01
PLT (10 ⁹ /L)	1025.60 ± 53.50	911.00 ± 47.85	826.56 ± 57.10	946.50 ± 57.56

Data are presented as means ± SEM. † $p < 0.05$ vs Sham-veh; # $p < 0.05$ vs MI-veh.

AXA, anti-factor Xa activity; PT, prothrombin time; APTT, activated partial thromboplastin time; WBC, white blood cell count; RBC, red blood cell count; HCT, hematocrit; HGB: hemoglobin; PLT, absolute automated platelet count.

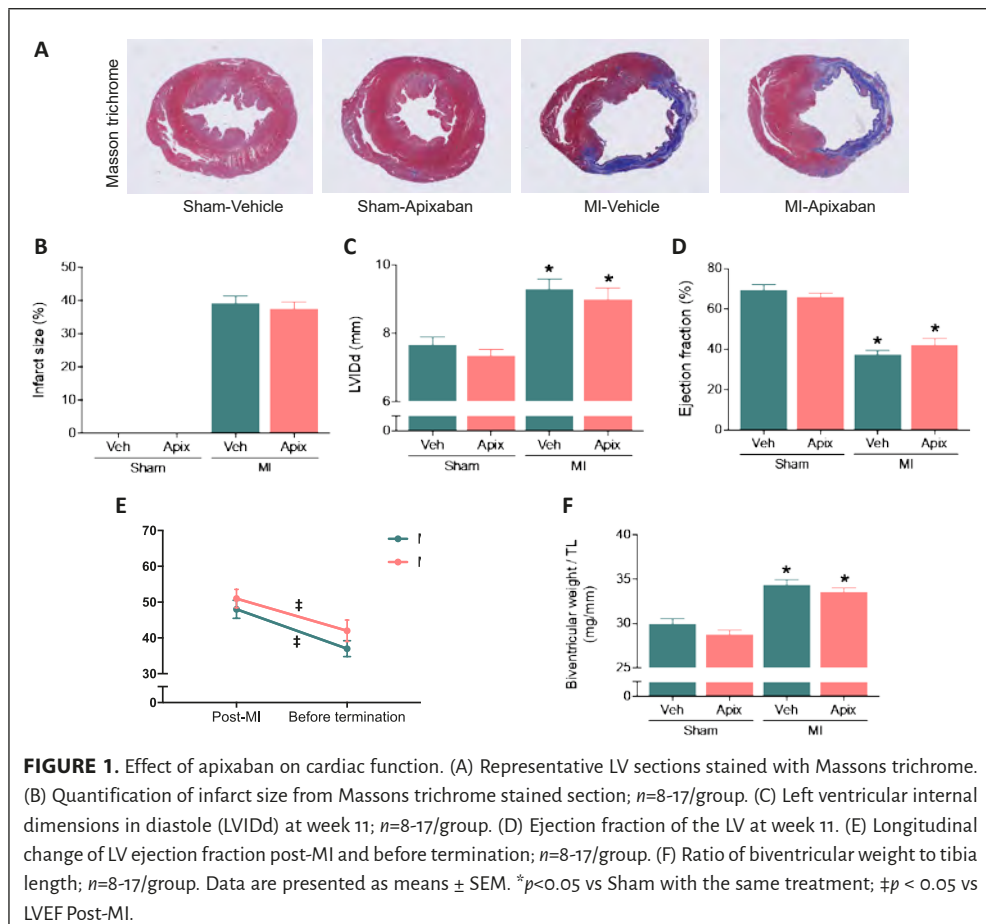
Efficacy and safety of apixaban in rats with HF after MI

Efficacy of apixaban

Daily food intake and water intake were comparable among the groups (Table 1). As expected, plasma PT and APTT were not affected by apixaban treatment (Table 1) [24]. In rats treated with apixaban, the plasma AXA activity was increased by fivefold as compared with that in vehicle-treated rats (Table 1).

Safety of apixaban

We did not observe any bleeding events during the study, nor did we observe reductions in haemoglobin levels (Table 1) or detect blood in the urine (Supplementary Figure S1). Furthermore, apixaban did not affect other haematological parameters such as white blood cells (WBC), red blood cells (RBC), haematocrit (HCT) and platelet (PLT) counts (Table 1). Body weight was similar in apixaban and vehicle treated groups (Table 1).



Effect of apixaban on cardiac function

The average MI size was $38 \pm 2\%$ and was comparable between MI-vehicle and MI-apixaban (Figure 1A and 1B). MI surgery resulted in cardiac dilatation and a marked reduction in LVEF (Figure 1B and 1C). At the initiation of therapy, two weeks after MI, all indices of left ventricular function were comparable between the MI-Apixaban and the MI-vehicle groups. As expected, a progressive deterioration in LVEF was observed in the MI-vehicle group over the 10-week treatment period (Figure 1E). Treatment with apixaban did not alter cardiac function and all echocardiographic parameters remained comparable between the MI-vehicle- and the MI-apixaban-treated animals (Figure 1, Table 2). All other relevant echocardiographic parameters at week 11 are depicted in Table 2. The infarcted rats were hemodynamically compromised, as reflected by a decrease in contractility and relaxation ($dP/dt \text{ max-min}$), and an increase in LVEDP. Treatment with apixaban did not alter these parameters (Table 3). Moreover, no significant difference was found in systolic blood pressure or diastolic blood pressure among the groups (Table 3).

TABLE 2. Echocardiography parameters in sham operated and post-myocardial infarction rats at week 11.

Parameters	Sham-veh	Sham-apixaban	MI-vehicle	MI-apixaban
IVSd (mm)	2.09 ± 0.15	2.46 ± 0.13	1.89 ± 0.10*	1.81 ± 0.13*
LVIDd (mm)	7.67 ± 0.22	7.34 ± 0.18	9.29 ± 0.31*	8.99 ± 0.33*
LVPWd (mm)	2.18 ± 0.13	2.23 ± 0.27	1.88 ± 0.12	1.85 ± 0.12
IVSs (mm)	3.45 ± 0.18	3.64 ± 0.27	2.39 ± 0.15*	2.31 ± 0.21*
LVIDs (mm)	4.52 ± 0.25	4.38 ± 0.22	7.52 ± 0.37*	7.04 ± 0.35*
LVPWs (mm)	3.14 ± 0.13	2.86 ± 0.34	2.37 ± 0.10*	2.42 ± 0.14
FS (%)	41.41 ± 2.37	37.40 ± 1.68	19.02 ± 1.22*	22.17 ± 1.76*
EF (%)	70 ± 2	66 ± 2	37 ± 2*	42 ± 3*

Data are presented as means ± SEM. * $p < 0.05$ vs Sham with the same treatment.

IVS, interventricular in diastole (d) and systole (s), respectively; LVID, left ventricular internal dimensions in both diastole (d) and systole (s); LVPW, the thickness of left ventricle posterior wall in diastole (d) and systole (s); FS, fractional shortening; EF, left ventricular ejection fraction.

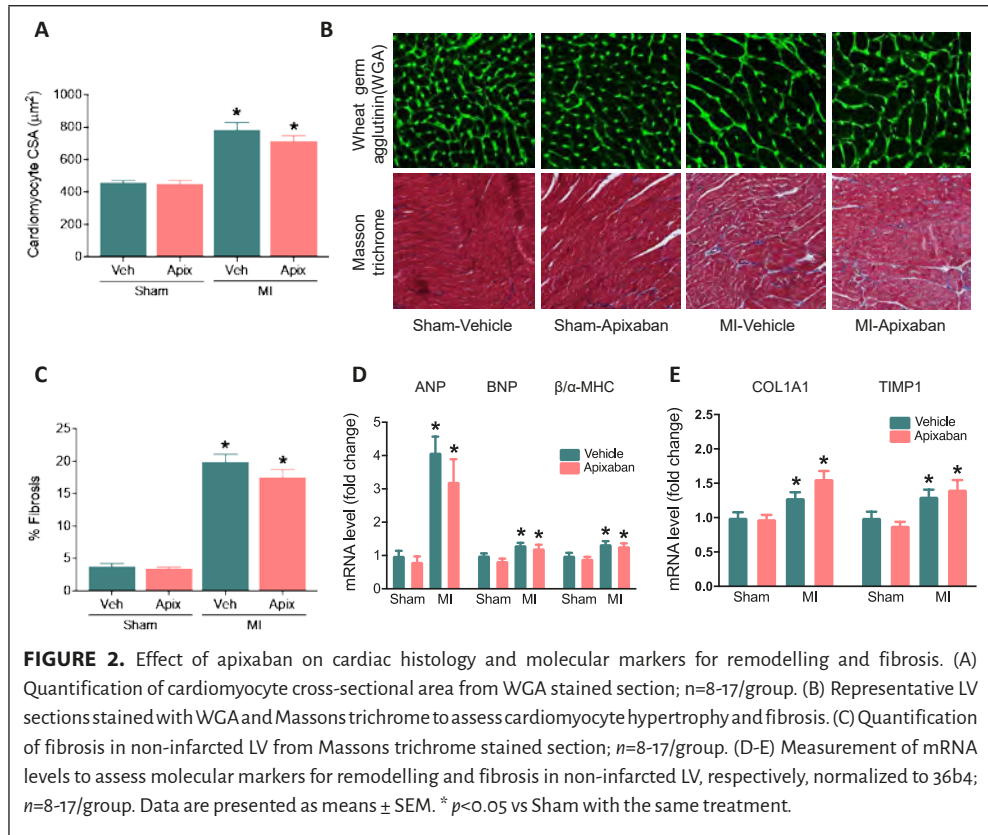
Effect of apixaban on cardiac histology and molecular markers for remodelling and fibrosis

The biventricular weight/tibia lengths ratio was calculated as a marker of hypertrophy and was found to be significantly increased in MI-vehicle rats compared with that in sham operated rats (*Figure 1E*). The MI rats demonstrated increased cardiomyocyte cross sectional-area and fibrosis compared with the control rats (*Figure 2A-C*). However, apixaban therapy did not affect the extent of cardiac hypertrophy nor did it influence the degree of LV fibrosis (*2A-C*).

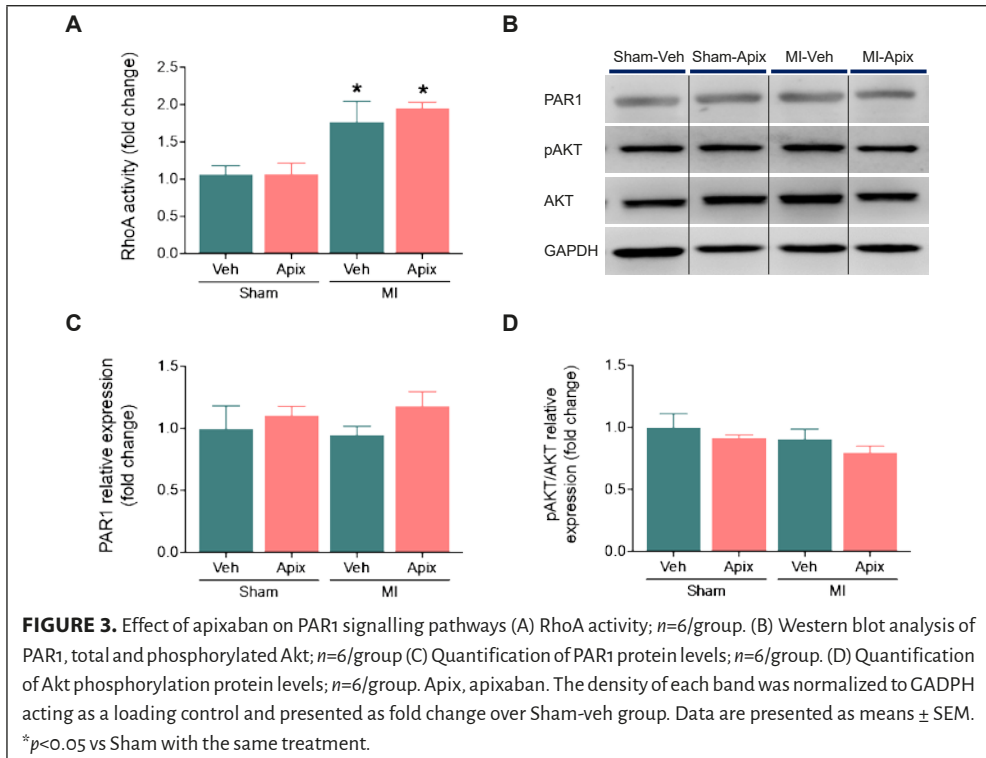
MI surgery increased the myocardial expression of atrial natriuretic peptide (ANP), brain natriuretic peptide (BNP) and increased the relative expression of foetal (β -MHC) compared to adult (α -MHC) myosin heavy chain isoform (i.e. β -MHC/ α -MHC ratio), as markers for foetal gene reprogramming in heart failure (*Figure 2D*). The mRNA levels of cardiac fibrosis markers collagen, type I, alpha 1 (COL1A1) and tissue inhibitor of metalloproteinases 1 (TIMP1) were also significantly increased in the hearts of rats following MI, compared to the hearts of sham rats (*Figure 2E*). Apixaban treatment had no effect on ANP, BNP, β -MHC/ α -MHC ratio, COL1A1 or TIMP1 mRNA levels (*Figure 2D and 2E*).

Effect of apixaban on PAR1 signalling pathways

Next we aimed to determine whether apixaban influenced the activity of thrombin-related pathways downstream of the PAR1 receptor. Binding of thrombin to the PAR1 receptor results in the activation of RhoA, which in turn phosphorylates Rho-associated coiled-coil kinase (ROCK) [25]. The RhoA/ROCK pathway has been shown to contribute to cardiac remodelling in HF [26,27]. It has also been reported that PAR1 activates AKT and ERK, two well established regulators of cardiac remodelling, independent from RhoA [16,28].



Next we determined the effects of apixaban on myocardial RhoA activity. As expected, RhoA activity was increased after MI, but the RhoA activity did not differ between the MI-vehicle and the MI-apixaban group (Figure 3A). Furthermore, apixaban did not influence the phosphorylation levels of AKT (Figure 3B and 3D), nor did it affect the activation of ERK1/2 (data not shown). Finally, the LV protein expression levels of the PAR1 receptor were comparable between all groups (Figure 3B and C), indicating that the results above could not be explained by aberrant expression of the PAR1 receptor.



DISCUSSION

We tested the hypothesis that treatment with FXa inhibitors apixaban would improve cardiac function and ameliorate cardiac remodelling in rats with HF after a large transmural anterior MI. For this purpose, we used a well-established model of chronic post-MI HF and ensured that the degree of LV dysfunction was similar in the apixaban and vehicle-treated groups at the initiation of therapy. Furthermore, we used a clinically relevant dose of apixaban that also appeared to be safe and effective, as evidenced by a consistent 5-fold increase in AXA and by the absence of (occult) bleeding. We demonstrate that treatment with apixaban did not influence LV function, nor did it influence LV dilatation or LV hypertrophy. Moreover, histological and molecular markers for pathological LV remodelling were also not influenced by apixaban and the activity of signal transduction pathways downstream of the PAR1 was unaltered. These findings suggest that the inhibition of FXa with a safe, effective and clinically relevant dose of apixaban does not influence cardiac remodelling in the chronic phase after MI. In addition, our findings suggest that the role of thrombin-mediated PAR1 receptor activation to the pathophysiology of HF is limited. Our findings are in line with the results from the COMMANDER HF trial and do not support the use of FXa inhibitor in HF patients with the aim to amend the severity of HF.

HF reflects a procoagulant state, because all prerequisites for thrombosis as described in Virchow's law are met: abnormalities in blood flow, in the vessel wall, and in blood constituents [29,30]. Aberrant platelet activation and increased levels of pro-coagulant factors reflect abnormalities in blood constituents. Impaired contractility and dilated chambers perturb myocardial blood flow and endothelial damage and reduced endothelium-dependent vasodilation reflect the changes in the vessel wall [31]. It has been previously recognized that patients with HF have an increased risk of thromboembolic events, both systemic and venous. The prothrombotic molecules, such as fibrinogen and von Willebrand factors, have been found to be elevated in subjects with HF. Moreover, previous studies showed that the thrombotic risk associated with HF appears to increase with the severity of the disease [32,33]. A large Danish prospective cohort study demonstrated that the risk of stroke and thromboembolic event was increased in patients with HF independent of the CHA₂DS₂-VASc score. Furthermore, patients with HF and a CHA₂DS₂-VASc score ≥ 4 , the absolute risk of thromboembolic events was even higher in patients without than with concomitant AF [34]. In addition, the prognosis of HF markedly deteriorates after a thromboembolic event occurs, suggesting that interventions to reduce thromboembolic events may improve prognosis in HF patients [35]. Another mechanism by which the hypercoagulable state could contribute to the progression of HF is coronary microvascular embolization leading to MI [36,37].

Anticoagulants are often prescribed in HF patients without AF that are considered to be at high risk for stroke, such as those with an LV thrombus. There is, however, very little evidence to support lenient prescription of anticoagulants in patients with HF and sinus rhythm [38]. In the Warfarin versus Aspirin in Reduced Cardiac Ejection Fraction (WARCEF) trial, warfarin did not result in a meaningful reduction in the rates of ischaemic stroke, intracerebral haemorrhage or death from any cause [39]. Furthermore, the effect of FXa inhibitor rivaroxaban on clinical outcomes was recently tested in the COMMANDER HF ("A Study to Assess the Effectiveness and Safety of Rivaroxaban in Reducing the Risk of Death, Myocardial Infarction, or Stroke in Participants with Heart Failure and Coronary Artery Disease Following an Episode of Decompensated Heart Failure") trial [40]. The authors randomized >5000 patients with chronic ischemic HF with reduced ejection fraction but without atrial fibrillation to a low dose of rivaroxaban or placebo. Rivaroxaban was safe and well tolerated, but did not affect the incidence of the combined endpoint of all-cause mortality, stroke or a myocardial infarction. In both the WARCEF and the COMMANDER HF study, anticoagulation did reduce the incidence of ischemic stroke, suggesting that the lack of effect was not dose related. Our results confirm and extend upon the results of the COMMANDER HF trial and provide mechanistic underpinnings that explain the neutral outcomes. Furthermore, our findings suggest that clinically relevant doses of FXa inhibitors do not influence PAR mediated signalling and cardiac remodelling.

To the best of our knowledge, our study is the first to study the effect of Xa inhibition on cardiac remodeling in rats with established LV dysfunction. Several studies have, however, evaluated the early effect FXa inhibition on myocardial ischemia/reperfusion injury and post-infarct remodelling. The indirect FXa inhibitor fondaparinux has been shown to reduce infarct size following 2 h of reperfusion in a rat model of myocardial ischemia-reperfusion [41]. The effects of FXa inhibitors are more variable as Flierl *et al.* did not observe infarct size reduction when rivaroxaban was given in rats with permanent coronary ligation [42]. However, a similar study by Bode *et al.* indicated that administration of rivaroxaban immediately after surgery resulted in reduction in infarct size and improvements in cardiac function. Conversely, when rivaroxaban was initiated 3 days after MI surgery, no effect on infarct size or cardiac function was observed [43]. Taken together, the available evidence on the effect of FXa inhibitors on cardiac function are in line with our observations and dispute their utility in chronic HF setting.

There are four PAR isoforms, but PAR1 and PAR2 are the predominant isoforms in the heart [13,16]. PAR1 is activated by thrombin and FXa, but PAR2 is only activated by FXa. Both receptors are expressed in cardiomyocytes and cardiac fibroblast. Cleavage of PARs results in activation of several G protein coupled receptors and their downstream signalling pathways, including RhoA/ROCK, the MAPK pathways, ERK 1/2 and ERK5 [16]. Several lines of evidence indicate that PARs are activated in HF and contribute to disease progression. First, PAR1 expression and the activity of its downstream signal transduction pathways are increased in murine models of chronic HF and in the ventricles of HF patients with ischemic or idiopathic dilated cardiomyopathy [14,15]. Second, the activation of PARs in cultured cardiomyocytes induces pathological hypertrophy, reflected by increases in cell size and sarcomeric organization, the activation of the foetal gene program and perturbations in cardiac calcium handling [28]. Third, PAR1 activation in cardiac fibroblasts induces a pro-fibrotic state reflected by enhanced proliferation and increased expression of transforming growth factor Beta (TGF- β) [13,28,44]. Fourth, PAR1 activation is strongly pro-inflammatory as it induces the expression of interleukin (IL)-6, IL-8, and monocyte chemoattractant protein (MCP-1) [12,13]. Yet, the most robust evidence comes from studies in PAR1-knockout (KO) mice and mice with overexpressing the PAR1. In a PAR1-KO mouse model reduced cardiac remodelling was observed after I/R injury; however, infarct size was not affected. Accordingly, mice with cardiomyocyte specific overexpression of PAR1 exhibited eccentric hypertrophy and dilated cardiomyopathy [16]. Additionally, PAR signalling activate RhoA/ROCK pathway [25,45] which has been shown to mediate fibrosis in the heart [26]. Protein levels of ROCKs as well as RhoA activity were significantly increased in CHF patients [27]. Furthermore, deletion of ROCK attenuates HF progression and improve the cardiac performance in mice [46,47]. These findings provided a clear rationale for our studies.

However, in contrast to our expectations we did not observe any changes in protein levels for PAR1 protein, nor did we detect differences in the activation of the AKT and ERK 1/2 signal transduction pathways downstream of PAR1 [48]. These findings are in consistence with a previous study [49] in which PAR1 protein expression did not differ at 12 weeks after MI induction or apixaban treatment. Moreover, it has been previously published that up-regulated AKT exerts cardioprotective effects in models of preconditioning resulting in limiting infarct size [50,51]. However, AKT is not regulated in a chronic ischemia setting[52], and this finding was also seen in our study. Our observation that ERK1/2 protein levels were also not affected in post-MI LV dysfunction is in accordance with similar studies in post-MI LV dysfunction [53]. Consistent with other reports, we were able to detect a clear increase in RhoA activity after MI, but this was not affected by treatment with apixaban.

Other studies have demonstrated evidence that FXa inhibitors can influence PAR receptor signalling. For instance, Bukowska *et al.* showed that FXa inhibitor rivaroxaban reduced MAP kinase activity and diminished the upregulation of PARs, ICAM-1, LOX-1, and IL-8 and in human atrial tissue cultures in media containing activated FXa [54]. Interestingly, FXa has also been shown to activate ERK1/2 and induce pro-inflammatory cytokines in alveolar epithelial cells, which was suppressed by FXa inhibitor edoxaban [55]. In addition, the FXa inhibitor rivaroxaban did inhibit cardiac FXa activity and reduced cardiac fibrosis in model of transverse aortic constriction [56]. Our results may therefore have been different in other disease models or if we would have induced a murine model prone to develop cardiac thrombi [57].

STUDY LIMITATIONS

Despite strengths related to the direct measures of cardiac structure, function and other haematological parameters, our study does have limitations. First, in accordance with the previous study [49], we did not observe changes in PAR1 protein levels in post-MI HF model. Furthermore, the activity of downstream signalling pathways of the PAR1 was not increased after MI and apixaban did not alter this. It is possible that the activation of PAR1 and other thrombin-related pathways is more pronounced in other models or settings. Second, we started the treatment 2 weeks after MI when infarct healing had completed, we cannot exclude that FXa inhibition could be beneficial during earlier stages of post-infarct remodelling. Third, we employed apixaban at a dose that was within the safety range (FDA application No. 202155Orig1s000). Based on the AXA levels, this dosage is comparable to a 5 mg dose in humans [17]. The outcome of our study may have been different when a higher dose had been used. The clinical relevance of a study with high-dose apixaban is, however, limited as anticoagulants have a narrow therapeutic window and the benefits are often offset by the associated increased risk of bleeding events [39]. Our study does not exclude

the possibility that thrombin-related pathways and PARs contribute to cardiac remodelling in HF. Our findings do, however, question the utility of FXa inhibitors as a pharmacological therapy to attenuate cardiac remodelling. Importantly, our study is in line with the neutral effects of the COMMANDER-HF trial, and the molecular insights do not hint towards a direct effect of apixaban on cardiac muscle structure and function.

CONCLUSIONS

FXa inhibition with apixaban does not influence pathological cardiac remodelling after a MI. These data do not support the use of FXa inhibitor in HF patients with the aim to amend the severity of HF.

CLINICAL PERSPECTIVES

It has been suggested that coagulation factors such as factor Xa (FXa) and thrombin promote maladaptive cardiac remodelling and could promote heart failure (HF) development via activation of the protease-activated receptors (PARs) in myocardial tissue. If this hypothesis is true, cardiac remodelling would be amendable by treatment with anticoagulants. To test this hypothesis, rats with HF after myocardial infarction (MI) were treated with the FXa inhibitor apixaban or a matching vehicle. While apixaban was effective in inhibiting FXa activity, it did not affect the activity of PAR1 signaling pathways, nor did it affect cardiac function and cardiac remodelling after MI. These findings are in line with the results of the recent COMMANDER HF trial, and do not support the use of FXa inhibitors in HF patients with the aim to improve heart function or to modulate the clinical course of HF.

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COMPLIANCE AND ETHICAL STANDARD

Ethics approval and consent to participate

The study was approved by the University of Groningen Ethical Committee on Animal Experimentation (IvD number: 16487-02-001). The investigation conforms to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health and in accordance with national regulations.

Competing interest

SRY, HHWS, KTN, MMD, DjvV and BDW do not report conflicts of interest relative to this report. The UMCG, which employs all authors, has received research grants and/or fees from AstraZeneca, Abbott, Bristol-Myers Squibb, Novartis, Roche, Trevena, and ThermoFisher GmbH. Dr. de Boer received personal fees from MandalMed Inc, Novartis, and Servier.

Disclaimer

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SUPPLEMENTARY MATERIAL

Supplementary Tables

TABLE S1. List of primers used in this study.

Transcript	Forward primer sequence	Reverse primer sequence
ANP	ATGGGCTCCTTCTCCATCAC	TCTACCGGCATCTTCTCCTC
BNP	ACAATCCACGATGCAGAAGCT	GGGCCTTGGTCCTTTGAGA
α -MHC	GACAACTCCTCCCGCTTTGG	AAGATCACCCGGGACTTCTC
β -MHC	TCTGGAGGCCTTTGGCAATG	GATGCCAACTTTCTGTGTC
COL1A1	ACAGCGTAGCCTACATGG	AAGTTCCGGTGTGACTCG
TIMP1	AGAGCCTCTGTGGATATGTC	CTCAGATTATGCCAGGGAAC
36B4	GTTGCCTCAGTGCCTCACTC	GCAGCCGCAAATGCAGATGG

TABLE S2. List of primary antibodies for Western blotting.

Antibody	Species/Clonality	Source (Catalog No.)
phosphorylated-Akt	Rabbit/Monoclonal	Cell signaling (#4060)
Akt	Rabbit/Monoclonal	Cell signaling (#4691)
PAR1	Mouse/Monoclonal	Santa Cruz (sc-13503)
GAPDH	Mouse/Monoclonal	Fitzgerald (10R-G109A)